

## Alteration of Ecdysteroid Titre by Thyroxine and Juvenile Hormone Analogue (Methoprene) in *Bombyx mori* (Lepidoptera: Bombycidae)

Pengaruh Tiroksin dan Analog Hormon Juvenil (Metopren) terhadap Pola Titer Ekdisteroid *Bombyx mori* (Lepidoptera: Bombycidae)

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### Abstrak

Penelitian ini dilakukan untuk mengetahui pengaruh hormon tiroksin dan analog hormon juvenil (metopren) terhadap pola titer hormon ekdisteroid hemolimf selama instar lima *Bombyx mori*. Instar dua *B. mori* diberi pakan daun murbei yang telah diberi tiroksin dengan konsentrasi 1500 ppm. Larva kemudian dibiarkan ganti kulit sampai instar tiga dan empat tanpa perlakuan. Tujuh puluh dua jam setelah ganti kulit pada instar lima, larva disuntik sebanyak 10 µl metopren dengan konsentrasi 15 atau 25 ppm. Titer ekdisteroid hemolimf dideteksi dengan Kromatografi Cair Kinerja Tinggi. Hasil yang diperoleh menunjukkan bahwa perlakuan dengan tiroksin ataupun kombinasi tiroksin-metopren menyebabkan terjadinya perubahan pola titer ekdisteroid dibandingkan dengan kontrol. Ekdisteroid terdeteksi dua hari lebih awal pada perlakuan tiroksin dan titer ekdisteroid pada puncak tertinggi lebih tinggi jika dibandingkan dengan kontrol. Kombinasi tiroksin-metopren tampaknya menyebabkan penekanan terhadap titer ekdisteroid. Hal ini terlihat dengan terjadinya penurunan titer ekdisteroid segera setelah penyuntikan dengan metopren sebelum mencapai titer puncak tertinggi.

**Kata kunci:** *Bombyx mori*, tiroksin, metopren, ekdisteroid, titer

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### Introduction

Application of hormones to *Bombyx mori* to improve the quality of silk has been suggested by many reports. For example, Thyagaraja *et al.*, (1991) reported that thyroxine could be used to shorten larval period and at the same time increased cocoon and silk weight up to 150% as compared to controls, while Akai *et al.*, (1985) reported that juvenile hormone analogue (methoprene) could be used to increase silk ratio up to 30% over control. Similar result was also reported by Miranda *et al.*, (2002) topical application of methoprene caused an increase in the weight of *B. mori* silk gland and cocoon.

Despite the fact that application of either thyroxine or juvenile hormone analogue can affect the cocoon and silk weight; we have decided to study the effects of the combination of the two substances, thyroxine and methoprene, to the larval period and silk gland and cocoon quality of *B. mori*.

Application of thyroxine would affect the insect's physiological processes. For example, treatment with thyroxine to *B. mori* larvae increased the ecdysteroid titre almost two-fold on larvae treated during first instar to fifth instar and 3.5-fold for larvae treated during second instar only as compared to untreated larvae. Treatment with thyroxine also increased the production of larvae haemolymph protein

(Thyagaraja *et al.*, 1991). Meanwhile, treatment with juvenile hormone analogue (Methoprene) would inhibit secretion of prothoracicotropic hormone and synthesis of ecdysteroid (Sakurai *et al.*, 1998).

During fifth instar of *B. mori* under normal conditions, ecdysteroid titre in haemolymph increased significantly on the fourth day and after that, it increased gradually and fluctuated each day (Sakurai *et al.*, 1998). The fluctuation was due to the existence of juvenile hormone.

Considering those facts, we thought it was necessary to study the effects of the applications of combination of thyroxine and methoprene on haemolymph ecdysteroid titre pattern during fifth instar of *B. mori*, in order to see if the combination of both hormones would cause an alteration in haemolymph ecdysteroid titre pattern during fifth instar of *B. mori* compared to that under the normal conditions.

## Materials and Method

### Experimental insects

*B. mori* type Polyhibrid bivoltine C.301 were reared under laboratory conditions in the School of Life Sciences and Technology, Bandung Institute of Technology, Indonesia. The animals were reared on mulberry leaves at 25-27°C, 85% RH and 12 h light: 12 h dark cycle.

### Preparation of thyroxine solution

Thyroxine was obtained from thyroid powder Grade II (Sigma Ltd. USA). 150 mg thyroxine was diluted in 5 ml sodium hydroxide 0.1 N (4 g NaOH + 1000 ml aquadest). The solution then was added by 95 ml aquadest to obtain a thyroxin solution with concentration of 1.5 mg/ml (1500 ppm) (Thyagaraja *et al.*, 1991).

### Preparation of methoprene solution

Methoprene was obtained from Altosid 1.3 G (Sandoz Crop). Altosid contained 1.3% methoprene as an active ingredient. In this experiment, the concentrations of methoprene used were 15 and 25 ppm. To obtain a 15 ppm methoprene solution, an amount of 1.15 gram

of Altosid was diluted in 1 liter of aquadest, while 1.92 gram of Altosid was needed to make a 25 ppm methoprene solution.

### Treatment of *Bombyx mori*

Thyroxine was given *ad libitum* during the second instar. The minced leaves were dipped in thyroxine solution for 3 minutes, then left to dry. Third and fourth instars were not treated with thyroxine. Then, 72 hours after molting to fifth instar, treatment using methoprene was applied. Each individual was injected with 10 µl of 15 or 25 ppm methoprene solutions on the segmental junction at the lateral part of the larva.

### Removal of haemolymph and isolation of ecdysteroid

50 µl of haemolymph was drained every day from each fifth instar larva that has been anesthetized using ether, through a cut on its proleg. The haemolymph sample was put into a 1.5 ml microtube and was added with 500 µl mix of acetone-ethanol (1:1). The mixture then was stirred and kept in a temperature of -20°C until further processing.

Ecdysteroid isolation was conducted using method from Beydon (1985). Haemolymph was extracted using acetone-ethanol (1:1) with ratio of 1:10, and then the mixture was centrifuged at 10,000 g for 10 minutes. The supernatant was removed and the remaining solution was re-extracted. Supernatants from the first and second centrifugation were mixed and evaporated, then 500 µl hexane-methanol 80% (1:1) was added. When hexane-methanol phase (marked with the formation of a borderline between hexane and methanol) has occurred, the methanol phase (lower part) that contained ecdysteroid was removed and evaporated. The product was diluted in methanol and filtered using "Sep-Pak® Cartridges" and analyzed by HPLC.

### Quantitative and qualitative analysis of ecdysteroid by HPLC

Ecdysteroid diluted in methanol was analyzed by HPLC type Kromatopak Shimadzu CR-7A plus. The type of column used was Shim-pack CLC-ODS C18 0.6 mm in diameter

and 15 cm in length. The mobile phase used was a mixture of methanol-water (9:1) mixed isocratically. The flow rate was 1 ml/minute and the wavelength used to detect ecdysteroid was 254 nm (Beydon, 1985).

Before conducting the analysis, first the mobile phase was filtrated using cellulose acetate membrane filter (PTFE) 0.45  $\mu\text{m}$ . Ecdysteroid standard used as a comparison was ecdyson (obtained from Sigma Ltd. USA). Ecdyson standard was diluted with methanol pro analysis. To obtain the required concentration, further dilution was needed.

Qualitative analysis was conducted by comparing retention time between standard ecdysteroid and the sample. If the sample contained compounds that had the same retention time with the standard ecdysteroid, then the compound was assumed to be ecdysteroid. Co injection of standard and sample were conducted to confirm the existence of ecdysteroid in the sample.

Quantitative analysis to determine ecdysteroid concentration was conducted by comparing area width of the sample to that of the standard on a standard calibration curve. The curve was made by plotting the area width of the ecdysteroid standard to its concentration.

## Results and Discussion

### Ecdysteroid titre in haemolymph of fifth instar of *Bombyx mori*

As can be seen in Figure 1, ecdysteroid was detected two days earlier in larvae which had been treated with thyroxine during the second instar, as compared to the control (untreated) larvae.

Ecdysteroid in control larvae was first detected on the sixth day with the concentration of 99.83  $\mu\text{g/ml}$  and increased until the seventh day (115.45  $\mu\text{g/ml}$ ). Ecdysteroid titre on the seventh day was the highest during the fifth instar and would decrease until larvae stopped eating and started making cocoon. The lowest titre was achieved on the eighth day (58.45  $\mu\text{g/ml}$ ), two days before larvae started making cocoon.

In larvae treated with thyroxine during the second instar, ecdysteroid first detected on the fourth day. After the fourth day, ecdysteroid titre in *B. mori* haemolymph would increase gradually until reaching the maximum titre on the sixth day. The maximum peak of the titre was 33.34% higher than that of the control. This peak was detected one day earlier in treated larvae compared to the control.

In *B. mori* larvae treated with thyroxine and methoprene, there was a suppressing pattern in the haemolymph ecdysteroid titre. Ecdysteroid titre drastically decreased shortly after application of 15 ppm methoprene, from 82.71 to 64.71  $\mu\text{g/ml}$ . Two days after the application, ecdysteroid titre had suffered a decrease then increased drastically and reached the maximum titre peak on the seventh day (three days after treatment). Meanwhile, in the combination of thyroxine and methoprene of higher concentration (1500/25 ppm), one day after the application the ecdysteroid titre was 81.72  $\mu\text{g/ml}$  and did not differ significantly to the ecdysteroid titre one day earlier. Two days after treatment, the ecdysteroid titre increased again, but much lower compared to that in larvae treated using methoprene 15 ppm. In comparison, in larvae without methoprene treatment (control and larvae treated with thyroxine only), the ecdysteroid titre kept increasing and did not show any decrease before reaching the maximum ecdysteroid titre.

The maximum peak of ecdysteroid titre in larvae treated using combination of thyroxine-methoprene 1500/15 ppm was about 32.14% or 1.32 times higher than that of the untreated larvae. In larvae treated using combination of thyroxine and higher concentration of methoprene (1500/25 ppm), the maximum titre peak did not differ much from that of the control. Generally, application of a higher concentration of methoprene (25 ppm) caused a lower and relatively more stable ecdysteroid titre during fifth instar than that of other treatments.

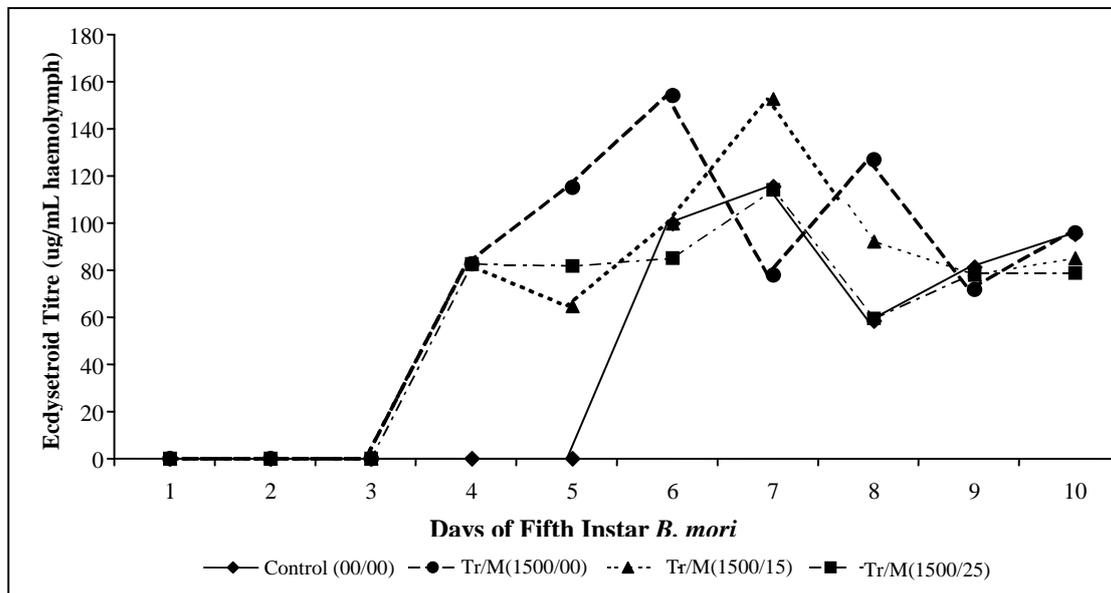
### Ecdysteroid titre in haemolymph of fifth instar of *Bombyx mori*

Thyagaraja *et al.*, (1991) suggested that *B. mori* feeding on thyroxine at earlier instar would eventually stimulate the brain to release

prothoracicotropin and regulate the prothoracic gland to release a precocious amount of ecdyson at the later instar. The present experiments seemed to substantiate their view as shown by comparison to the controls, in larvae treated with thyroxine during their second instars, ecdysteroid was detected two days earlier with titre of 82.71%, and had a maximum titre peak of 33.34% or 1.33 times higher.

Interestingly, applications of methoprene to fifth instar of *B. mori* seemed to cause a change in ecdysteroid biosynthesis. This hypothesis is supported by Mu and Leblanc (2004 in Tuberty & McKenney, 2005) who reported that exposure of JHAs caused a reduction in ecdysteroid gene expression, and it was possibly caused by competitive binding of the JHAs to the receptors required for regulating ecdysteroid production. In this research, it could be seen, for example, by a drastic decrease in ecdysteroid titre shortly

after methoprene applications, from 82.71 to 64.71  $\mu\text{g/ml}$ , during treatment with combination of thyroxine and methoprene 15 ppm. In *B. mori* larvae treated with a higher concentration of methoprene (25 ppm), the average of ecdysteroid titre was lower and relatively more stable than that of other treatments. The results of this study were generally almost identical to the results reported by Watson *et al.*, (1987) that the application of (7S)-hydroprone, an analogue to juvenile hormone, to *Manduca sexta* larvae would stimulate the prothoracic gland to produce ecdyson. This result also supported by Tuberty and McKenney (2005) who reported that exposure to juvenile hormone analogues affect ecdysteroid concentrations *in vivo* in crabs and shrimps larvae. The exposure of JHAs caused fluctuations in ecdysteroid concentrations throughout their larval stages.



**Figure 1.** Effect of thyroxine feeding and methoprene injection on ecdysteroid titre during the last larval instars. Control without hormone treatment (00/00); treatment with thyroxine (Tr) 1500 ppm given during second instar (1500/00); combination of thyroxine 1500 ppm with methoprene (M) 15 ppm (1500/15); combination of thyroxine 1500 ppm with methoprene 25 ppm (1500/25). Quantity of ecdysteroid was determined using HPLC and was calibrated using ecdyson as a standard of comparison. Day one (0 hours) was the first day after molting to fifth instar

In the present experiment, at first, it seemed that the drastic decrease in ecdysteroid titre occurred after applications of a lower concentration of methoprene (15 ppm), even though the titre would rapidly increase again in a large amount. The stronger effect of thyroxine-methoprene combination with lower concentration than that of the higher concentration shortly after applications of methoprene (one day after application) was probably due to the characteristics of most hormones. Hormones have optimum dose/concentration; therefore, increasing the amount of hormones will not necessarily increase the performance.

Under normal conditions, the production of juvenile hormone titre by the corpora allata in the middle of the fifth instar of *B. mori* is supposed to decrease or stop. But, because of the application of methoprene during stadium of 72 hours, the titre of juvenile hormone inside the blood remained high. It led to a change on the normal condition of ecdysteroid hormone inside the insect's body.

Ecdysteroid titre that decreased shortly after an application of methoprene would bounce back and reach the maximum titre peak on the seventh day (three days after the treatment). It was probably because methoprene in haemolymph had been decreased or totally vanished. The same phenomenon was reported by Trivedy *et al.*, (1995) in which fifth instar of *B. mori* treated with a low dose of juvenile hormone R394 topically to stadium of 24 hours did not show any distinct effect to its ecdysteroid titre until the sixth day. On the seventh day, ecdysteroid titre dropped significantly until it began making a cocoon. Larvae treated with low dose at a stadium of 48, 72, and 96 hours showed a reduction in their ecdysteroid titre starting from the fifth day to the seventh day and would increase again on the eighth day, when they began making cocoon.

The maximum titre peak of haemolymph ecdysteroid in control larvae obtained in this experiment on the seventh day did not differ with the result of the study conducted by Trivedy *et al.*, (1995) and Gu & Chow (1996). Ecdysteroid titre increased from 3 ng/ml on the first day to 70 ng/ml on the second day, then

was sustained on the level between 70 to 136 ng/ml during the second and fifth day, while the highest titre peak occurred on the seventh day (Gu & Chow, 1996). But, it was different from the result of the research by Thyagaraja *et al.*, (1991) where the highest titre peak of the haemolymph ecdysteroid of control was measured on the eleventh day of the fifth instar of *B. mori*. While in larvae treated with thyroxine, the highest titre peak of the ecdysteroid occurred on the tenth day. It is possible that the difference was probably due to a difference in the strain of silkworm used and environmental factors.

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